Amendments to the Specification:

Kindly delete paragraph 1of page 1 with the following new paragraph:

--This application is a continuing application of U.S. Application Serial No. 09/581,013 filed July 7, 2000, now issued as U.S. Pat. No. 6,733,983, which is a §371 national stage application of international PCT Application NO. PCT/NL98/00701 (WO 99/30162). This application asserts priority of European Patent Office Application Serial No. 97203851.7 filed on December 8, 1997. The specifications of U.S. Application Serial No. 09/581,013, now issued as U.S. Pat. No. 6,733,983, PCT/NL98/00701 and European Patent Application Serial No. 97203851.7 are incorporated herein by reference.--

Please insert the following new section at page 4, after line 15, and before line 16:

-- Brief Description of Drawings

FIG. 1A shows a positive reaction in a particle agglutination test with saliva using KP90 and polystyrene particles;

FIG. 1B shows a negative reaction under the same conditions as in FIG. 1A.--

At page 10, in Example 1, please delete the sentence from lines 17-19 and in its place insert the following:

-- KP90 was prepared from starting material made from crude mycobacterial mass as follows:

The mycobacteria were cultured in commercially available Sauton medium supplemented with 2 g $MgSO_4$, 8 g citric acid, 2g K_2HPO_4, 16 g asparagine, 2 g (Fe^+) ammonium citrate, 240 ml glycerol. The bacteria were cultured under standard conditions. The cells were harvested by filtration of the culture medium with a 12 μm filter. The cells were subsequently resuspended in 20 ml PBS (phosphate-buffered salt solution) (pH 7.4) and the harvested cells were autoclaved

under a pressure of 15 Psi for 20 minutes in order to deactivate and sterilize the bacteria. The thus obtained bacterial mass can be stored at -80°C.

In order to determine the quantity of starting material a 1/100 dilution of the harvested autoclaved suspension in PBS was made. The optical density thereof, measured at 420 nm ($O.D._{420}$) must be 0.1. If necessary the concentrated bacterial mass is supplemented with PBS (pH 7.4) until the correct O.D. is obtained. An $O.D._{420}$ of 0.1 indicates the presence of $7x10^{11}$ bacteria per 30 ml, which is equivalent to 12 g. wet weight of the bacterial mass.

For preparation of a crude mycobacterial extract 5 g wet weight of the bacterial mass was washed three times with PBS (pH 7.4). Centrifuging was then carried out at 3000 x g until the mass precipitated. The pellet was suspended in 50 ml PBS and stirred carefully to reduce formation of lumps to a minimum. To prevent lump forming 0.05% Tween 80 was optionally added. In order to avoid bacterial contamination 3 mg penicillin/streptomycin was added to this solution. The concentration was then brought with PBS to 2 g wet weight/ml.

The bacterial mass was subsequently broken open using an automatic French-X-press or RIBI press (American Instruments Company, Trevenollab. Inc. Maryland). The buckets were pre-cooled overnight at -20°C. Before use, the buckets were held in a mixture of ethanol and dry ice (-20°C). After the buckets were filled with 1 g bacterial mass per bucket of 5 ml and cooled at -80°C for 20 minutes, the buckets were placed in the French-X press and 12 tons of pressure were applied by pushing in the plunger of the press. The buckets were then removed and cooled again at -80°C for 20 minutes. The buckets were inverted and treated for the second time. 10 tons of pressure were applied the second time. Cooling and breaking were then repeated a number of times, normally about 5 times. The disrupted cells were eluted with a suitable volume of PBS and subsequently centrifuged at 4°C at 300 x g for 10 minutes in order to remove the unbroken bacteria with the sediment. The collected supernatant was then centrifuged at 4°C and. 145,000 x g for 2 hours. The pellet was suspended in 0.1 M Tris-HCl (pH 7.2), 0.01 M EDTA which contained 20 mM $MgSO_4.7H20$ in a concentration of about 1 g per 10 ml. 1 mg RNase and 1 mg DNase were added per 10 ml volume. Incubation then took place overnight at 4°C with careful stirring. Incubation thereafter took place for 1 hour at 37°C and the lysate was

centrifuged at 300 x g and 4°C for 10 minutes in order to remove the last-remaining unbroken bacteria (this is further referred to as "starting material").--